

Localization of the amino acid substitution site in a fast migrating variant of human serum albumin

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Albumin M₁/Fg is an Italian genetic variant of human serum albumin arising from a Lys → Glu substitution which has been located in a CNBr fragment (CNBr VII) corresponding to the -COOH terminal portion of the molecule [(1984) *J Chromatogr* 298, 336–344]. Tryptic peptides of CNBr VII from normal and M₁/Fg albumin have been purified by reverse-phase high-performance liquid chromatography (RP-HPLC) and submitted to comparative structural studies. The amino acid sequence of the tryptic peptide of M₁/Fg variant that differs from the corresponding fragment of the normal serum albumin shows that the Lys → Glu substitution responsible for this variant is located at position 573. This region of the albumin molecule is involved in the binding of long chain fatty acids.

Human serum albumin Genetic variant Tryptic peptide HPLC Amino acid sequence Point mutation

1. INTRODUCTION

Genetic variants of human serum albumin have been identified on the basis of their different electrophoretic mobility which can be either faster or slower than that of normal serum albumin [1,2]. Although a very large number of variants have been reported [1–3], little information has been obtained, so far, about the structural basis of their genetically determined variation. Only two variants have been characterized with respect to amino acid sequence: albumin Oliphant in which glutamic acid 570 is replaced by lysine [4], and albumin Mexico 2 that rises from replacement of aspartic acid 550 by glycine [5].

Albumin M₁/Fg is a fast variant which has been found in one homozygous and several heterozygous subjects within the same Italian family: it is similar in electrophoretic mobility to the Gent albumin, but different from the Naskapi fast variant which has never been found in Italy [6]. We have previously shown [7] that a Lys → Glu substitution which occurs in the COOH terminal portion of the molecule (residues 549–585), is responsible for the M₁/Fg fast variant. The present

results state that Lys 573 is replaced by glutamic acid.

2. EXPERIMENTAL

2.1. Sample preparation

Human normal serum albumin and M₁/Fg variant have been isolated from serum of a normal subject and, respectively, of a 15-year-old boy homozygous for M₁/Fg albumin, according to Winter et al. [4], and comparatively processed. Carboxymethylation, cyanogen bromide cleavage, HPLC separation of CNBr fragments and amino acid analysis of the two proteins have been carried out as in [7]. CNBr VII from normal and M₁/Fg albumins have been submitted to tryptic digestion, according to Swenson et al. [8]: after 16 h at room temperature, the digest was brought to pH 2 by addition of trifluoroacetic acid (TFA).

2.2. LC apparatus

Reverse-phase HPLC was developed on a Waters Associates (Milford, MA, USA) liquid chromatograph. The system consisted of two M6000 pumps, a U5K sample injector, a model 680

automated gradient controller and a variable wavelength detector (Japan Spectroscopic Co., Tokyo). The separation was carried out on a μ -Bondapak C-18 column, 10 μ m particle size, 30 cm \times 3.9 mm (Waters Associates).

2.3. Procedure

Sample (200 μ l), corresponding to 50 nmol tryptic digest, was separated on the μ -Bondapak C-18 column equilibrated with 0.05% aqueous trifluoroacetic acid (solvent A). Elution of peptides was achieved by use of a 60 min linear gradient from 0 to 50% of CH₃CN containing 0.05% TFA (solvent B) at a flow rate of 2 ml/min. All runs were performed at room temperature; individual peaks were collected manually and vacuum-dried. Peptides eluted under each peak have been identified by amino acid and N-terminus analyses, and are designated by arabic numerals starting from the N-terminal of the CNBr VII frag-

ment. Amino acid analysis was performed according to Dévényi [9]; N-terminal amino acid was identified as the dansyl derivative [10]

3 RESULTS AND DISCUSSION

Amino acid analyses of CNBr fragments allowed us to state that the amino acid substitution responsible for Mi/Fg variant is located in CNBr VII, corresponding to residues 549–585 of the albumin sequence: amino acid composition reported in table 1 shows that CNBr VII from Mi/Fg variant contains a glutamyl instead of a lysyl residue.

The chromatographic patterns of tryptic peptides obtained from normal and Mi/Fg CNBr VII are given in figs 1,2. Comparison of the elution profiles shows the absence of T₄ + T₅ peptide in Mi/Fg CNBr VII tryptic digest. The T₄ + T₅ peptide obtained from normal albumin CNBr VII

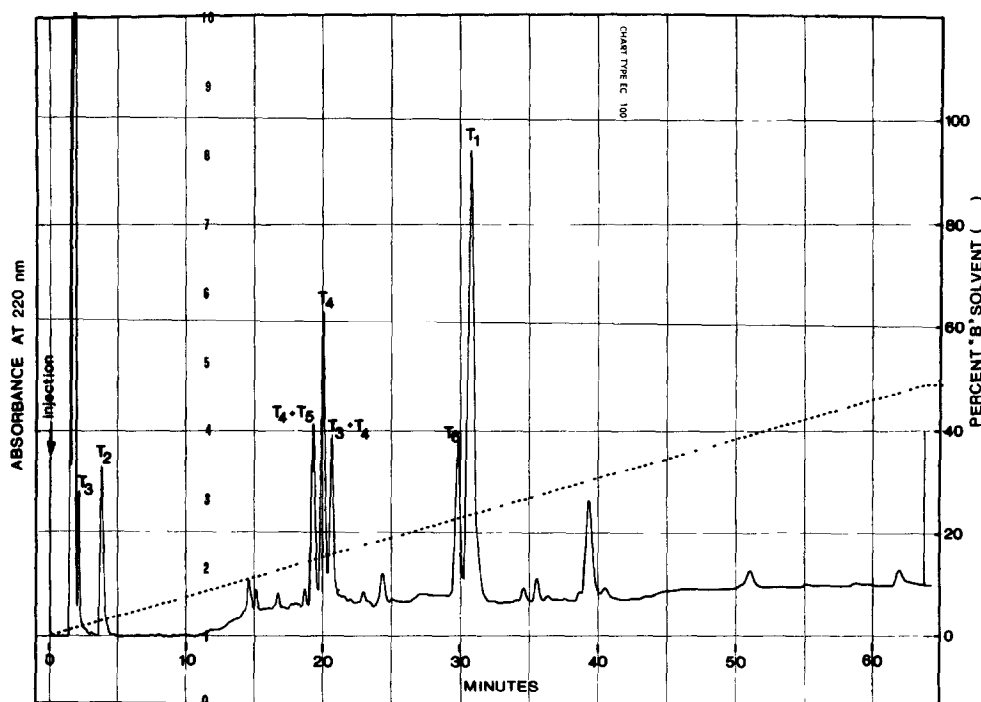


Fig.1. Elution pattern of tryptic digest of CNBr VII fragment obtained from normal human serum albumin. The mixture was dissolved in 0.05% aqueous TFA, pH 2 (solvent A) and 200 μ l (corresponding to about 50 nmol digest) was injected onto a μ -Bondapak C-18 column (30 cm \times 3.9 mm), equilibrated with solvent A. The elution was performed at room temperature and a 60-min linear gradient from 0 to 50% acetonitrile containing 0.05% TFA (solvent B) was employed as indicated. Flow rate: 2 ml/min. Absorbance range: 0.64 full scale

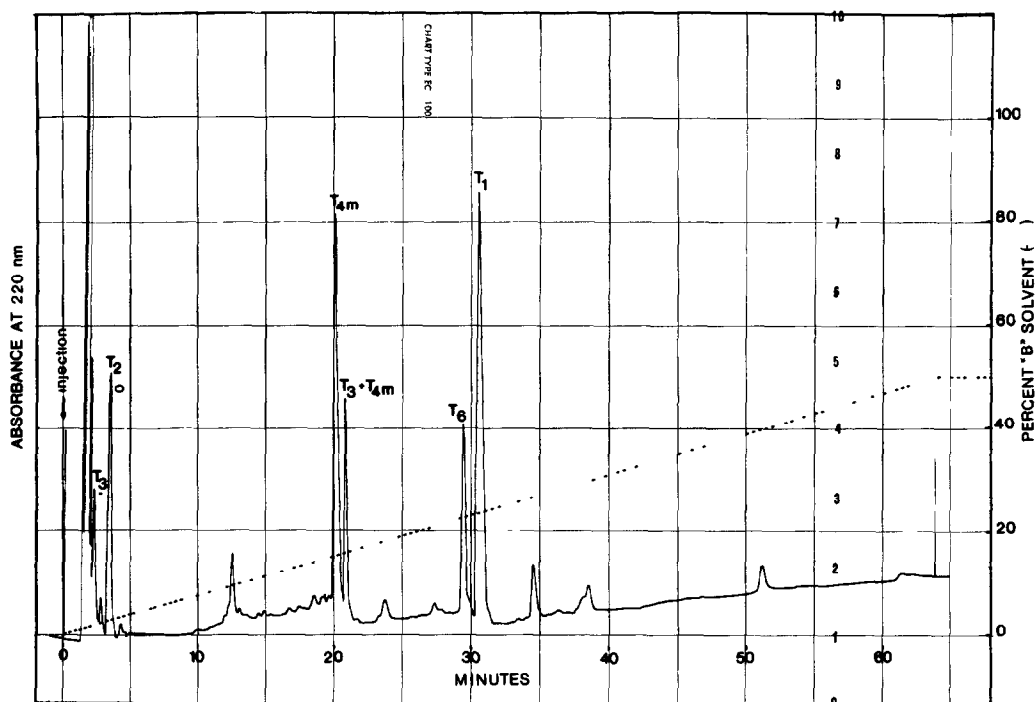


Fig.2. Elution pattern of tryptic digest of CNBr VII fragment from M1/Fg variant obtained under the conditions used in fig.1

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REFERENCES

- [1] Weitkamp, L.R., Salzano, F.M., Neel, J.V., Porta, F., Geerdink, R.A. and Tarnoky, A.L. (1973) *Ann. Hum. Genet.* 36, 381-392
- [2] Tarnoky, A.L. (1980) *Adv. Clin. Chem.* 21, 101-146
- [3] Schell, L.M. and Blumberg, B.S. (1977) in: *Albumin Structure, Function and Uses* (Rosenoer, V.M., Oratz, M. and Rothschild, M.A. eds) pp.113-141, Pergamon Press, New York.
- [4] Winter, W.P., Weitkamp, L.R. and Rucknagel, D.L. (1972) *Biochemistry* 11, 889-896
- [5] Franklin, S.G., Wolf, S.I., Zweidler, A. and Blumberg, B.S. (1980) *Proc Natl Acad Sci USA* 77, 2505-2509
- [6] Vanzetti, G., Porta, F., Prencipe, L., Scherini, A. and Fraccaro, M. (1979) *Hum. Genet.* 46, 5-9.
- [7] Iadarola, P., Ferri, G., Galliano, M., Minchiotti, L. and Zapponi, M.C. (1984) *J. Chromatogr.* 298, 336-344
- [8] Swenson, R.P., Williams, C.H. jr, Massey, V., Ronchi, S., Minchiotti, L., Galliano, M. and Curti, B. (1982) *J. Biol. Chem.* 257, 8817-8823.
- [9] Dévényi, T. (1968) *Acta Biochim. Biophys. Acad. Sci. Hung.* 3, 429-431
- [10] Hartley, B.S. (1970) *Biochem. J.* 119, 805-822
- [11] Gray, W.R. (1972) *Methods Enzymol.* 25, 333-344
- [12] Dugaiczky, A., Law, A. and Dennison, O.E. (1982) *Proc. Natl. Acad. Sci. USA* 79, 71-75
- [13] Dayhoff, M.O., Hunt, L.T., McLaughlin, P.J. and Jones, D.D. (1972) in: *Atlas of Protein Sequence and Structure* (Dayhoff, M.O. ed.) vol.5, pp.17-30, Natl. Biomed. Res. Found., Silver Spring, MD, USA.
- [14] Berde, G.B., Hudson, B.S., Simoni, R.D. and Sklar, L.A. (1979) *J. Biol. Chem.* 254, 391-400.
- [15] Shaklai, N., Garlick, R.L. and Bunn, H.F. (1984) *J. Biol. Chem.* 259, 3812-3817